



Review Article



Research Progress on Leptin in Metabolic Dysfunction-associated Fatty Liver Disease

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Abstract

Metabolic dysfunction-associated fatty liver disease (MAFLD) poses a significant challenge in modern medicine due to its high prevalence. The pathogenesis of MAFLD involves a complex dysmetabolic process consistent with the “multiple-hit” hypothesis. This process includes excessive triglyceride (TC) accumulation within hepatocytes, lipotoxicity, insulin resistance (IR), chronic low-grade inflammation, and increased oxidative stress. The role of leptin in the liver has been extensively studied, demonstrating both direct effects on hepatic cells and indirect actions mediated through the central nervous system (CNS). In MAFLD, leptin modulates several physiological processes: it improves glucose metabolism by enhancing insulin sensitivity and lowering glucose levels; regulates lipid metabolism by promoting β -oxidation and TC export while inhibiting lipogenesis; and contributes to fibrogenesis by upregulating transforming growth factor- β (TGF- β) expression and activating hepatic stellate cells (HSCs) and the immune response. This review explores the structure of leptin, its primary physiological functions, its potential role in MAFLD pathogenesis, and its promise as a novel therapeutic target.

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Introduction

Metabolic dysfunction-associated fatty liver disease (MAFLD), the recently proposed term replacing nonalcoholic fatty liver disease according to a multi-society Delphi consensus statement, has emerged as the most common chronic liver disease,^{1,2} affecting over 25% of the general adult population and more than 50% of individuals with type 2 diabetes mellitus (T2DM).^{3,4} MAFLD is diagnosed when fatty liver is detected through histology, imaging, or blood biomarkers

in individuals with one of the following: overweight/obesity (ethnic-specific), T2DM, or metabolic dysregulation with two or more risk factors.⁵ Approximately 10–30% of individuals with MAFLD progress to metabolic dysfunction-associated steatohepatitis (MASH), a more advanced stage characterized by hepatic inflammation and hepatocellular injury (ballooning).⁶ MAFLD and MASH can lead to severe outcomes such as cirrhosis and hepatocellular carcinoma (HCC), and are also associated with extrahepatic complications, particularly atherosclerotic cardiovascular disease and ischemic stroke, which represent the leading causes of death in this population.^{7–9} Moreover, the global syndemic framework, highlighted in The Lancet Commission Report, emphasizes the interconnectedness of metabolic disease, cardiovascular disease, disability, cancer, and premature death, all of which share common biological mechanisms and societal determinants.¹⁰ Beyond genetic predisposition, major risk factors for MAFLD include obesity and/or sarcopenia, insulin resistance (IR), and metabolic comorbidities such as dyslipidemia and T2DM.^{11–14} MAFLD is recognized as an independent condition, meaning that its diagnosis does not exclude the contribution of other factors to liver dysfunction.¹⁵ Both extrahepatic factors—such as adipokine production, caloric and nutrient imbalance, and IR—and intrahepatic mechanisms—including impaired fatty acid oxidation, mitochondrial dysfunction, endoplasmic reticulum stress, oxidative stress, and activation of resident macrophages—drive the onset of MAFLD and its progression to MASH.¹⁶ Understanding the pathophysiology of MAFLD and identifying molecular targets for diagnosis and treatment remain crucial research priorities.

Leptin, a prominent adipokine primarily secreted by adipocytes, plays a central role in appetite regulation and energy homeostasis.¹⁷ It acts by stimulating anorexigenic pathways and suppressing orexigenic pathways within the central nervous system (CNS).¹⁸ Beyond appetite control, leptin regulates multiple physiological processes, including lipid and glucose metabolism and immune responses.¹⁸ Insights into leptin's role in hepatic metabolism largely stem from studies using ob/ob (leptin-deficient) and db/db (leptin receptor-deficient) mice. These models exhibit hepatic IR, TC and lipid accumulation, steatosis, and inflammation—phenotypes that are partially reversed by leptin administration.^{19,20} Although leptin facilitates hepatic lipid clearance and mobilization, leptin resistance, commonly seen in obesity, may limit its effectiveness in reducing steatosis.^{19,20} Furthermore, elevated circulating leptin levels, frequently observed in obese individ-

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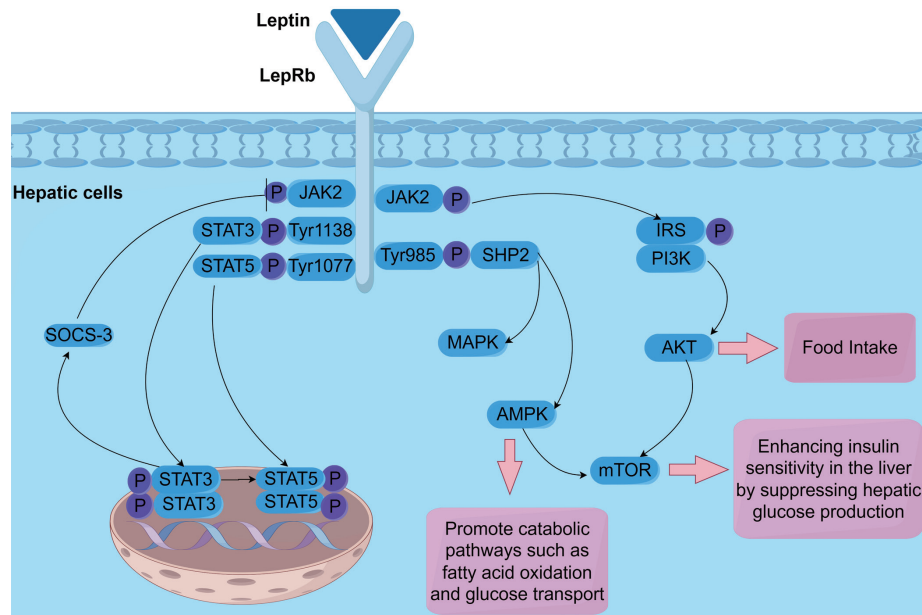


Fig. 1. Signal transduction of the leptin receptor in hepatic cells. Akt, protein kinase B; AMPK, 5'-adenosine monophosphate-activated protein kinase; ERK, extracellular signal-regulated kinase; Grb2, growth factor receptor-bound protein 2; IRS, insulin receptor substrate; JAK, Janus kinase; MAPK, mitogen-activated protein kinase; PI3K, phosphatidylinositol 3-kinase; PTP, tyrosine-protein phosphatase; SHP2, tyrosine-protein phosphatase 2 containing SH2; STAT, signal transducer and activator of transcription; SOCS3, suppressor of cytokine signaling 3.

uials, are linked to hepatic inflammation and fibrosis, further implicating leptin in MAFLD progression.

This review provides a comprehensive overview of the interplay between leptin and MAFLD. We summarize experimental and clinical evidence linking leptin to MAFLD pathogenesis and evaluate its therapeutic potential, with a focus on its regulatory roles in IR, lipid metabolism, inflammation, and oxidative stress.

Leptin: Structure and receptor

The discovery of leptin began in the early 1950s with the identification of the ob/ob mouse, a model characterized by severe obesity and hyperphagia.²¹ In the 1960s, the db/db mouse, another hyperphagic and obese diabetic model, was described.²² In 1994, Jeffrey Friedman's team successfully cloned the obese gene and named the protein it encoded leptin.²³ The following year, the leptin receptor (Ob-R) was cloned and found to be mutated in the db/db mouse.²⁴ Leptin is a polypeptide hormone composed of 167 amino acids, transcribed from the human OB gene located on chromosome 7, which contains three exons and two introns.²³ Structurally, leptin exhibits the three-dimensional features of a four-helix bundle cytokine. It includes a short signal peptide (21 amino acids) and a longer functional segment of 146 amino acids. Although predominantly synthesized and secreted by subcutaneous white adipose tissue, leptin is also produced in several peripheral tissues and in the CNS.²⁵ Leptin secretion is positively correlated with adipocyte size and is regulated by multiple factors, including body fat mass, metabolic state, circadian rhythm, and hormones such as insulin, glucocorticoids, and leptin itself.^{26–28} While classically regarded as a marker of long-term energy stores, leptin has pleiotropic functions: it modulates immune and inflammatory responses, neuroendocrine axes, autonomic nervous activity, cardiovascular function, reproduction, angiogenesis, osteogenesis, and hematopoiesis.^{29–34} In addition, leptin influenc-

es gluconeogenesis, insulin sensitivity, and lipid and carbohydrate metabolism in the liver; skeletal muscle regeneration; lipolysis in adipose tissue; and nutrient utilization in the small intestine by regulating mucus and hormone secretion, nutrient absorption, gastric emptying, and intestinal motility.³⁵

Leptin exerts its effects by binding to its cell surface receptor, Ob-R, encoded by the Ob-R gene in rodents. Throughout this discussion, murine nomenclature and numbering are used.^{24,36–38} Alternative splicing of the Ob-R transcript produces several isoforms. These include the signaling-competent Ob-Rb (LepRb), which has a long intracellular tail; shorter isoforms with truncated intracellular domains (Ob-Ra, Ob-Rc, Ob-Rd); and a secreted form consisting of only the extracellular domain (Ob-Re).^{39,40} CNS-expressed Ob-R isoforms are critical for energy balance, metabolism, feedback regulation, and immune function.⁴¹ Ob-Rb is also expressed in peripheral tissues such as skeletal muscle, adipose tissue, liver, and pancreatic β -cells, suggesting autocrine and paracrine roles in energy regulation.^{42,43} Shorter isoforms, Ob-Ra and Ob-Rc, are involved in leptin transport across the blood-brain barrier.⁴⁴ Leptin levels are inversely associated with the binding and clearance functions of these soluble receptors.^{45,46} However, only the long Ob-Rb isoform has a complete intracellular domain capable of transmitting signals upon ligand binding.⁴⁷

Leptin receptor signaling and MAFLD

Upon leptin binding, Ob-Rb in hepatocytes activates intracellular signaling via JAK2 (Janus kinase 2) phosphorylation (Fig. 1). JAK2 then phosphorylates three tyrosine residues (Tyr1077, Tyr1138, and Tyr985) on the intracellular domain of Ob-Rb.²⁰ Tyr1077 activates STAT5 (Signal transducer and activator of transcription 5), while Tyr1138 activates both STAT5 and STAT3 (Signal transducer and activator of transcription 3). Tyr985 engages the SHP2/MAPK (Tyrosine-protein phosphatase 2 containing SH2/ Mitogen-activated protein kinase)

cascade.^{48–50} Activated STAT3 upregulates SOCS-3 (Cytokine signaling 3 suppressor), a negative feedback regulator of leptin and insulin signaling.^{32,51,52} Elevated SOCS-3 contributes to hormone resistance, suggesting that SOCS-3 inhibition could represent a therapeutic approach in liver disease. In addition, IRS1/IRS2 (Insulin receptor substrate) phosphorylation and PI3K (Phosphatidylinositol 3-kinase) activation modulate JAK2 activity, playing a central role in leptin's regulation of food intake. Leptin also stimulates AMPK (5'-adenosine monophosphate-activated protein kinase) activity in peripheral tissues, promoting glucose uptake and fatty acid oxidation, while suppressing AMPK activity in the brain to regulate appetite.⁵¹ The PI3K/Akt (protein kinase B) pathway, which regulates mTOR signaling, is also activated and improves insulin sensitivity by reducing hepatic glucose output.⁵³

Leptin resistance

For leptin to exert its physiological effects, circulating concentrations must remain within a functional range. Excessive leptin, however, can induce receptor desensitization—even in genetically leptin-deficient mice.⁵⁴ In obesity, leptin resistance is characterized by impaired central and/or peripheral responsiveness despite elevated leptin levels.³⁵ This may result from impaired receptor signaling or defective transport across the blood-brain barrier (BBB).⁵⁵ Proposed mechanisms include suppression of the JAK-STAT pathway, downregulation of receptor expression, impaired post-receptor signaling, decreased histone deacetylase 5 activity in the hypothalamus, elevated C-reactive protein, endoplasmic reticulum stress, and hypothalamic inflammation.⁵⁶ SOCS3 plays a central role in diet-induced obesity by inhibiting STAT3 and downstream Ob-Rb signaling.^{57,58}

Leptin resistance increases the risk of diet-induced obesity and creates a feedback loop in which elevated leptin levels further exacerbate resistance. This resistance can manifest in the CNS and peripheral tissues and is referred to as central and peripheral leptin resistance, respectively.⁵⁹ Peripheral leptin resistance occurs in tissues such as skeletal muscle, adipose tissue, and the liver; it diminishes leptin's effectiveness in regulating lipid and carbohydrate metabolism and is closely associated with inflammation.^{60,61} Both leptin deficiency and leptin resistance are associated with hepatic lipid droplet accumulation, lymphocyte infiltration, and disturbances in glucose homeostasis.⁶² Disruptions in leptin signaling are also significantly associated with IR in MASH.⁶³ In humans, leptin resistance is influenced by obesity, lipodystrophy, and genetic variations, including Ob-R polymorphisms linked to MAFLD.^{64,65} Typically, obesity induces hyperleptinemia, which promotes inflammation and leptin resistance, leading to sustained postprandial hyperinsulinemia and ultimately metabolic dysfunction.

Possible models of leptin function in MAFLD pathogenesis

Glucose levels and IR

Leptin regulates blood glucose levels both directly, through peripheral tissues, and indirectly, via the CNS (Fig. 2).⁶⁶ In the CNS, these effects are mediated by leptin receptors (Ob-Rs) expressed on neurons.⁶⁷ Leptin primarily modulates glucose homeostasis in the context of obesity and IR through pro-opiomelanocortin-expressing neurons in the hypothalamic arcuate nucleus.^{68,69} Notably, intracerebroventricular leptin administration in animal models produces minimal changes in circulating leptin levels but improves IR and glycemic con-

trol, supporting the hypothesis that leptin primarily regulates glucose metabolism via central mechanisms, independently of food intake or body weight.⁷⁰ Peripherally, leptin regulates glucose levels by binding to receptors in multiple tissues and modulating pancreatic hormone secretion, lowering both insulin and glucagon.⁷¹ Although several studies show that leptin enhances glucose uptake, glycogenesis, and glucose oxidation in skeletal muscle, results have not been consistent across all investigations.^{72–74} Leptin reduces glucose uptake in adipocytes and decreases fatty acid and glycerol release, thereby limiting gluconeogenic substrate availability to the liver.⁷⁵ In the liver, leptin inhibits gluconeogenesis via IRS-2 and reduces hepatic TC content.^{76,77} Leptin may also interfere with insulin action by altering adipocyte sensitivity to lipid accumulation, reducing insulin receptor binding in the liver, and suppressing insulin secretion from pancreatic islets.^{45,78,79} However, human studies suggest leptin does not directly affect insulin secretion from pancreatic β -cells.⁸⁰ Serum leptin levels are not consistently associated with insulin or glucose concentrations, nor with the homeostasis model assessment of IR.⁸¹ Instead, leptin may lower blood glucose by reducing gluconeogenesis through substrate limitation. Furthermore, leptin-induced weight loss and reduced fat mass may improve insulin sensitivity, decrease glucose levels, and limit carbohydrate and lipid flux to the liver,⁸⁰ thereby suppressing *de novo* lipogenesis and reducing hepatic fat accumulation.⁸²

Hyperleptinemia, however, can damage pancreatic β -cells and impair JAK2/PI3K signaling in obese individuals with T2DM and MAFLD. Leptin resistance may further contribute to hyperinsulinemia and worsening IR, thereby increasing the risk of T2DM.⁸³ IR inhibits lipid oxidation while promoting triglyceride and fatty acid synthesis.^{45,78,84} Consequently, the leptin-insulin signaling axis that normally regulates glucose metabolism becomes disrupted. Hyperleptinemia also upregulates hepatic sterol regulatory element-binding protein 1 (SREBP-1), promoting lipogenesis.⁸⁵ Along with other pro-inflammatory adipokines, leptin has been strongly implicated in the development of IR, particularly in MAFLD.^{86,87}

Lipid metabolism

Leptin plays a dual role in fatty liver disease (Fig. 2). In early MAFLD, leptin reduces hepatic lipid accumulation and promotes lipid oxidation.^{19,88} Rodent models with impaired leptin signaling—including leptin-deficient *ob/ob* and *db/db* mice—support these findings.^{89,90} Hepatic steatosis arises from disruptions in both glucose and lipid metabolism, and leptin limits TC storage in adipose and non-adipose tissues, including the liver, to prevent lipotoxicity.²⁰ Leptin reduces hepatic lipid accumulation through vagal signaling by activating the JAK2-STAT3/AMPK pathway, independent of food intake.⁹¹ At physiological levels, leptin exerts anti-steatotic effects by suppressing lipogenesis and hepatic glucose production, thereby improving insulin sensitivity.^{52,92} These mechanisms explain the prevention or reversal of hepatic steatosis in *ob/ob* mice treated with leptin.⁹³ Leptin's anti-steatotic properties have also been demonstrated in non-obese mice with uncontrolled T1DM, in which leptin therapy reduced lipogenic and cholesterologenic transcription factors, lowering plasma and tissue lipid levels.⁹⁴ Leptin regulates hepatic lipid synthesis by modulating key transcription factors such as carbohydrate-responsive element-binding protein.⁹⁵ It may act synergistically with insulin and inhibit the production of very low-density lipoproteins (VLDL).^{94,96,97} Additionally, leptin may enhance VLDL-TC export, further reducing steatosis.⁹⁸ Some studies even suggest that leptin improves steatosis and IR in lipodystrophic mice.⁹⁹ Hackl *et al.* also reported leptin's protective role against ectopic lipid accumulation in the brain, underscor-

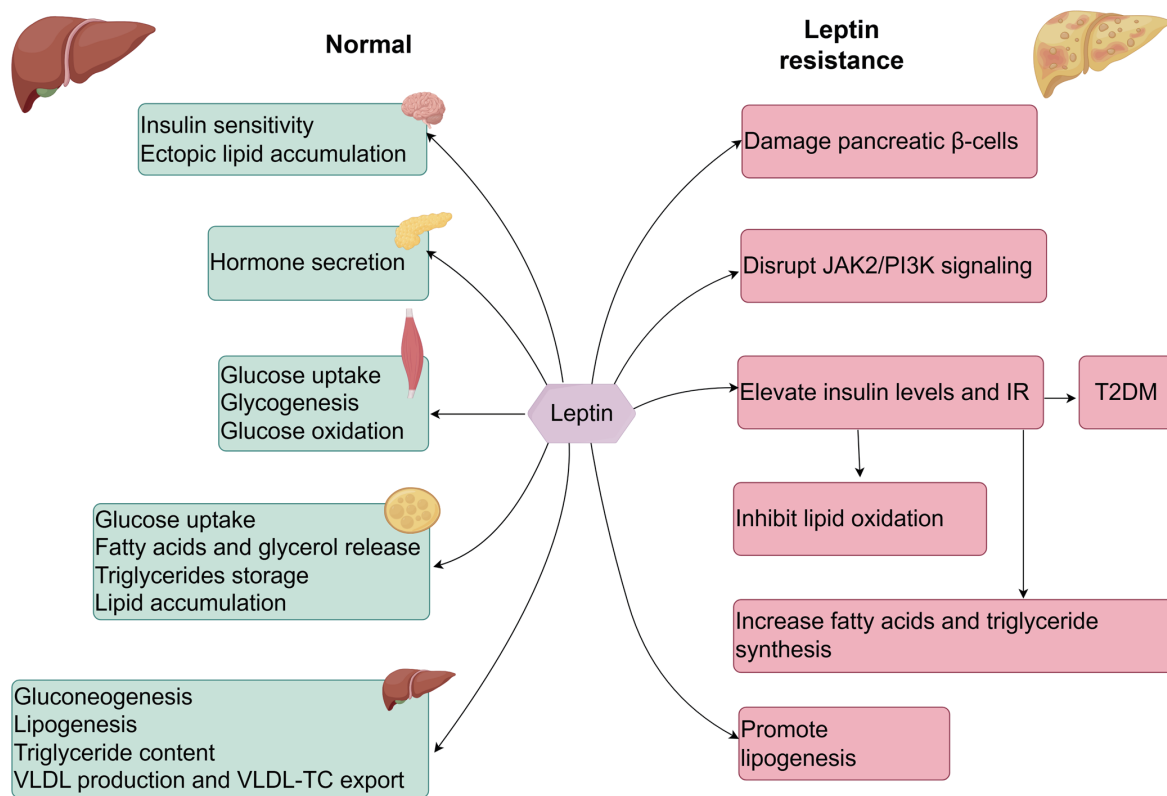


Fig. 2. Mechanisms of insulin resistance and lipid metabolism of leptin in MAFLD. In the early stages of MAFLD, leptin helps regulate glucose homeostasis, reduce hepatic lipid accumulation, and promote lipid oxidation. However, hyperleptinemia can damage pancreatic β -cells and disrupt JAK2/PI3K signaling, leading to elevated insulin levels and worsening IR. This IR inhibits lipid oxidation while promoting triglyceride and fatty acid synthesis. VLDL, very low-density lipoproteins; TC, Triglyceride; JAK, Janus kinase; PI3K, phosphatidylinositol 3-kinase; IR, insulin resistance; T2DM, diabetes mellitus, type2; MAFLD, metabolic dysfunction-associated fatty liver disease.

ing its therapeutic potential in obesity-related steatosis.¹⁰⁰

Despite these beneficial effects, elevated leptin levels often fail to resolve hepatic steatosis because of leptin resistance, a central feature of MAFLD pathogenesis.^{52,101,102} Mechanisms such as Tyr985 phosphorylation on Ob-Rb and increased SOCS-3 expression impair leptin signaling, particularly in the arcuate nucleus.¹⁰³ Leptin levels are positively correlated with steatosis severity, especially in individuals with high body mass index (BMI). In lean MAFLD patients, however, genetic and metabolic disorders (e.g., hypobetalipoproteinemia, celiac disease, cystic fibrosis) may play a greater role than leptin levels.¹⁰⁴ Leptin may also promote hepatic IR, attenuating its own anti-steatotic effects.¹⁰⁵ Cernea *et al.* reported increased MAFLD-related steatosis in T2DM patients,⁴⁵ while Pavlidis *et al.* found higher leptin levels associated with more severe steatosis in chronic hepatitis C.¹⁰⁶ Eshraghian *et al.* were the first to demonstrate that changes in leptin, adiponectin, and IR were associated with hepatic steatosis in liver transplant recipients.¹⁰⁷

Inflammation

The severity of hepatic steatosis in MAFLD correlates with progressive liver damage, ranging from simple steatosis to MASH (Fig. 3). While most patients present with isolated steatosis, approximately one-third progress to MASH, increasing the risk of advanced stages of MAFLD.¹ This progression is driven by inflammation, which arises when hepatic TC levels exceed the liver's adaptive capacity, resulting in lipotoxicity. Lipotoxicity is characterized by the production

of reactive oxygen species (ROS), endoplasmic reticulum stress, and hepatocellular injury.¹⁶ These cellular insults activate immune and apoptotic pathways, ultimately leading to cell death and contributing to fibrosis and cirrhosis over time.¹⁰⁸ Although leptin possesses anti-steatotic properties, it also exhibits pro-inflammatory and fibrogenic effects.¹⁰⁹ In animal models fed a high-fat diet, elevated leptin levels have been associated with inflammation and MASH.¹¹⁰ In the context of diet-induced obesity, leptin contributes to IR, T2DM, and chronic inflammation.^{111–113} Studies have shown that myeloid cells lacking leptin signaling demonstrate improved glucose tolerance in obese mice, underscoring leptin's role in low-grade systemic inflammation.¹¹⁴ Furthermore, leptin resistance has been linked to low-grade inflammation and steatosis in obese individuals, implicating leptin in the transition from simple steatosis to MASH.¹¹⁵

Early studies in ob/ob and db/db mice revealed immune deficiencies, including impaired antibody production, reduced cytotoxic activity, and increased susceptibility to autoimmune and allergic diseases.^{116,117} Leptin enhances lipopolysaccharide-induced production of tumor necrosis factor- α (TNF- α) in monocytes and macrophages.^{118,119} It can also independently stimulate inflammation by promoting the M1 macrophage phenotype.¹²⁰ Leptin deficiency exacerbates inflammatory cell infiltration and promotes MASH development.¹²¹ In murine preadipocytes and adipose-derived stromal cells, leptin facilitates lipid droplet formation and upregulates adipogenic and lipogenic signaling pathways, particularly through PPAR γ and SREBP-1c activation. Leptin

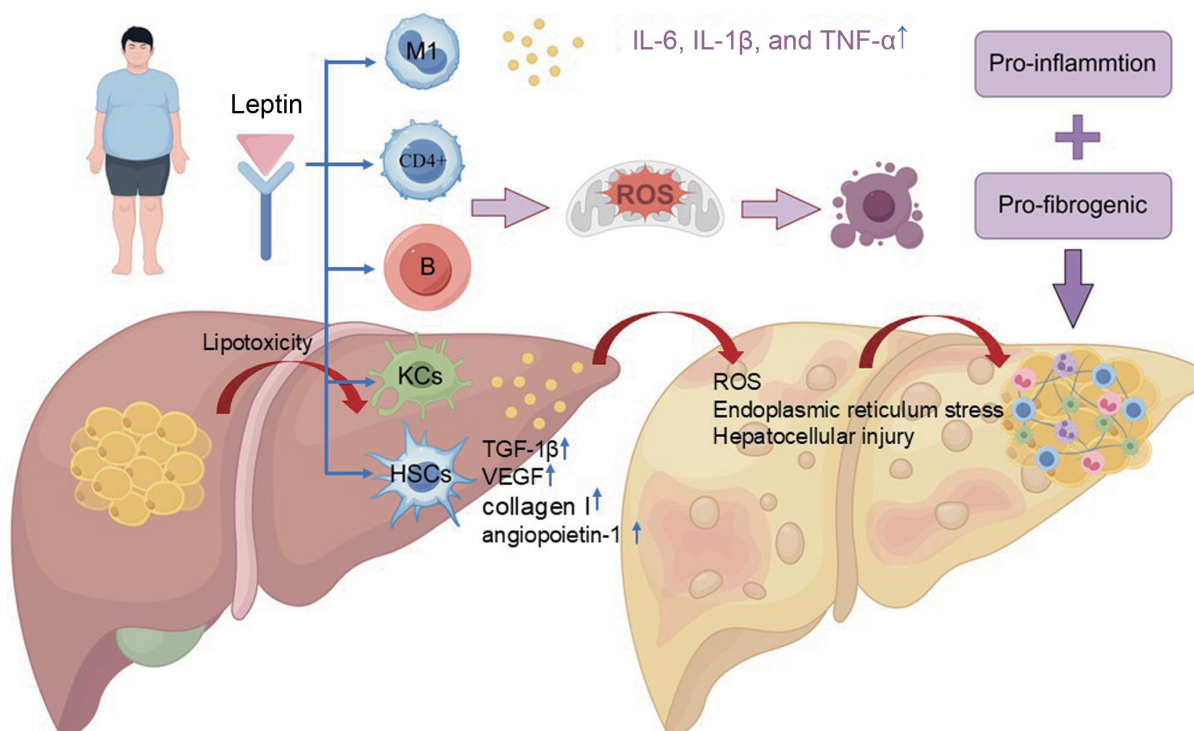


Fig. 3. Mechanisms of inflammation and oxidative stress of leptin in MAFLD. Leptin can independently stimulate inflammation by promoting the M1 macrophage phenotype while increasing pro-inflammatory cytokines such as IL-6, IL-1 β , and TNF- α . Leptin supports pro-inflammatory CD4⁺ T-cell and B-cell proliferation while inhibiting regulatory T-cell (Treg) expansion. These lymphocytes can release granzymes that produce ROS in mitochondria and activate caspases, ultimately inducing pyroptosis. It can also induce ROS formation in both phagocytic and non-phagocytic cells, including endothelial cells, cardiomyocytes, and HSCs. IL-6, interleukin-6; IL-1 β , interleukin-1 β ; TNF- α , tumor necrosis factor- α ; ROS, reactive oxygen species; M1, macrophage phenotype 1; KCs, Kupffer cells; HSCs, hepatic stellate cells; TGF- β 1, transforming growth factor- β 1; VEGF, vascular endothelial growth factor; MAFLD, metabolic dysfunction-associated fatty liver disease; $\uparrow$, increase.

also reduces interleukin (IL)-10 production while increasing pro-inflammatory cytokines such as IL-6, IL-1 β , and TNF- α .¹²² Conversely, TNF- α and IL-1 stimulate leptin secretion.^{123,124} Leptin supports pro-inflammatory CD4⁺ T-cell proliferation and inhibits regulatory T-cell expansion.^{125,126} It also plays a role in TH17 cell differentiation in human CD4⁺ T cells.¹²⁷ In B cells, leptin has been shown to promote the release of inflammatory mediators and accelerate immunosenescence. B cells from young, lean individuals treated with leptin displayed inflammatory and aging markers typically observed in B cells from individuals with obesity or advanced age.¹²⁸ In steatotic livers with active inflammation, OB-R expression is significantly increased in activated mouse CD4⁺CD8⁺ T cells and B cells. Leptin signaling promotes lymphocyte survival and function.¹²⁹ These lymphocytes can release granzymes that generate ROS in mitochondria and activate caspases, ultimately inducing pyroptosis.¹³⁰

Tissue inflammation frequently leads to additional tissue injury. Leptin levels are elevated in patients with liver fibrosis, particularly among those with high BMI, whereas leaner individuals exhibit reduced inflammation and fibrosis.¹⁰⁴ Some studies suggest that leptin receptors on Kupffer cells (KCs) and sinusoidal endothelial cells contribute to the expression of matrix-remodeling enzymes, thereby initiating fibrosis through hepatic stellate cells (HSCs) activation.^{89,104,131} Leptin also induces vascular endothelial growth factor expression in HSCs, which may drive irreversible cirrhosis and MASH progression.¹³² In KCs, leptin upregulates transforming growth factor- β , likely activating HSCs through paracrine signaling.^{89,104,131} Activated HSCs exacerbate hepatic inflammation and fibrosis by secreting

angiopoietin-1, collagen I, transforming growth factor- β 1 (TGF- β 1), and vascular endothelial growth factor (VEGF). These cells also produce leptin and express ObRb, forming a feedback loop that sustains HSCs proliferation and inhibits apoptosis, thereby amplifying hepatic inflammation and fibrosis.⁸⁹ Moreover, leptin can activate KCs through oxidative stress mediated by peroxynitrite.¹³³ This activation promotes the presence of CD8⁺CD57⁺ T cells, which are implicated in MASH progression.¹³⁴ Prolonged hyperleptinemia may stimulate HSCs, KCs, and sinusoidal endothelial cells, triggering both pro-inflammatory and pro-fibrogenic signaling cascades.⁵²

Oxidative stress

KCs are a major source of ROS in the liver, primarily through the activity of NADPH oxidase, the key enzyme responsible for ROS generation (Fig. 3).¹³⁵ Danger-associated molecular patterns, such as ATP (Adenosine Triphosphate), activate KCs and stimulate ROS production.¹³⁶ Additionally, lipid peroxidation products like 4-hydroxynonenal can activate HSCs,¹³⁷ which also produce ROS via phagocytic activity and NADPH oxidase expression.¹³⁸ The expression of cytochrome P450 2E1, an enzyme involved in fatty acid oxidation, is elevated in models of alcoholic steatohepatitis and MASH, contributing to oxidative liver damage.¹³⁹ Increased oxidative stress also promotes auto-oxidation of excess cholesterol, forming oxysterols. Elevated oxysterol levels have been observed in biopsy-proven MAFLD patients and are associated with pro-inflammatory, pro-apoptotic, and pro-fibrogenic effects.¹⁴⁰ A comparative study found that oxidative stress-related cel-

lular damage in MAFLD aligns with the multiple parallel hits hypothesis.¹⁴¹ In this study, involving seven control subjects, 23 patients with MAFLD, and 17 with MASH, levels of 8-hydroxy-2'-deoxyguanosine and 4-hydroxynonenal were elevated in the MAFLD and MASH groups, indicating oxidative DNA damage and lipid peroxidation. Furthermore, compared with a non-hyperphagic rat model, obese hyperphagic rats showed increased hepatic oxidative stress.^{16,142}

Watson *et al.* demonstrated that leptin treatment restored altered glutathione peroxidase levels in ob/ob mice, suggesting leptin's role in regulating antioxidant enzyme activity.¹⁴³ *In vitro* studies have shown that leptin decreases malondialdehyde and ROS levels while increasing glutathione content.¹⁴⁴ Additionally, leptin enhances natural antioxidant enzyme activity and suppresses inflammatory factor expression in 3T3-L1 preadipocytes, thereby reducing oxidative stress-induced cellular injury.¹⁴⁵ Leptin supplementation has also been effective in mitigating oxidative stress induced by a high-fat diet.¹⁴⁶ In one study, offspring of obese dams given oral leptin during the suckling period demonstrated greater antioxidant capacity and reduced inflammatory markers in the liver, retroperitoneal white adipose tissue, and plasma compared to those given vehicle control.¹⁴⁷ Leptin-treated male offspring also exhibited increased plasma adiponectin and a higher adiponectin/leptin ratio, which enhances the liver's anti-inflammatory and antioxidant functions.¹⁴⁸ The absence of leptin in pigs led to reduced JAK2-STAT3 and AMPK phosphorylation, resulting in increased fatty acid β -oxidation and mitochondrial autophagy—both contributing to oxidative stress in liver cells.¹⁴⁹ In human hepatoma cell lines, leptin treatment of ethanol-exposed cells reduced ROS generation.¹⁵⁰ Furthermore, leptin was shown to suppress oxidative stress responses in blood and reduce endotoxemia-induced rises in pro-inflammatory cytokines such as IL-1, IL-6, and TNF- α .¹⁵¹

However, leptin can also induce ROS formation in both phagocytic and non-phagocytic cells, including endothelial cells, cardiomyocytes, and HSCs.^{152–154} Elevated leptin levels have been linked to increased ROS production, primarily through NADPH oxidase activation.¹⁵⁵ Hyperinsulinemia and hyperleptinemia are considered key contributors to oxidative stress in individuals with obesity or T2DM.^{156,157} Systemically elevated leptin levels exacerbate oxidative stress and weaken antioxidant defenses, thereby intensifying hepatic inflammation, especially in alcoholic liver disease.¹⁵⁸ In mouse models with steatohepatitic lesions, high circulating leptin levels aggravated liver damage via NADPH oxidase activation, inducible nitric oxide synthase induction, and increased TNF- α and monocyte chemoattractant protein-1 release from KCs, all mediated through peroxynitrite-dependent pathways.¹³³ Leptin promotes oxidative and inflammatory effects through three primary mechanisms: leptin-induced protein radical formation, tyrosine nitration, and KCs activation.¹³³ It also upregulates CD14 expression on KCs, a receptor for the bacterial endotoxin lipopolysaccharide.¹⁵⁹ This upregulation heightens cellular sensitivity to harmful stimuli and enhances oxidative stress. Notably, CD14 overexpression has been implicated in the progression of steatohepatitis and liver fibrosis, even in the absence of prior steatosis.¹⁶⁰

Leptin in MAFLD: Evidence from clinical studies

Basal leptin levels correlate with BMI and reflect the body's nutritional status. In severe obesity, the physiological set point for weight regulation may shift to a higher leptin threshold.¹¹⁵ When BMI decreases to below 25 kg/m², fasting leptin levels fall by approximately 15 ng/dL, likely due to improved leptin delivery to the CNS and enhanced access to specific

neuronal populations.^{130,161} Low circulating leptin levels and increased leptin sensitivity, both of which exert anti-steatotic effects, are associated with several interventions, including physical activity,¹⁶² bariatric surgery,¹⁶³ dietary polyphenol intake,^{164,165} caloric restriction, and sustained weight loss.^{166,167} Leptin thresholds vary among individuals due to genetic factors,¹⁶¹ sex-related differences,^{115,168} and, more recently, the use of GLP-1 receptor agonists.¹⁶⁹ Individuals with a history of obesity may retain elevated leptin levels even after returning to normal weight.¹⁷⁰

Importantly, elevated leptin levels have been linked to MAFLD even in lean individuals and in the absence of IR.¹⁷¹ Multiple studies demonstrate that high leptin levels are associated with both the presence and severity of MAFLD in children and adults, suggesting that early-onset leptin resistance may contribute to disease pathogenesis.^{172–177} For instance, Marques *et al.* reported elevated leptin levels in MAFLD patients compared to healthy controls, with no significant difference between obese and non-obese groups—indicating that obesity is not a confounding factor.¹⁷⁸ Moreover, reducing leptin levels with antibodies has been shown to restore leptin sensitivity, suggesting a potential therapeutic strategy for obesity and diabetes.¹⁷⁹ These findings imply that early intervention to lower leptin levels in children could help prevent MAFLD.¹⁷² Previous studies also reported associations between elevated leptin and hepatic fibrosis in MAFLD. In one study, high serum leptin was identified as a significant risk factor for hepatic steatosis, and receiver operating characteristic analysis confirmed leptin as an independent predictor of liver fat accumulation.¹⁸⁰

However, not all findings are consistent. Canbakan *et al.* found that leptin was not an independent predictor of MAFLD or fibrosis severity; their study reported higher leptin levels in patients with moderate fibrosis compared to those with advanced fibrosis.⁸¹ Similarly, in a pediatric cohort, children with prepubertal obesity and fatty liver disease had significantly lower leptin z-scores (adjusted for BMI) than peers with normal liver ultrasound findings,¹⁸¹ with these z-scores inversely correlated with fatty liver severity. In adults, normal-weight Caucasians with MAFLD, commonly referred to as lean MAFLD, showed significantly lower circulating leptin levels than obese MAFLD patients, though no difference was observed between lean MAFLD patients and lean healthy controls.¹⁸² A meta-analysis reported higher leptin levels in MAFLD patients with fibrosis compared to non-MAFLD controls; however, this association weakened after excluding one study involving morbidly obese individuals.¹⁸³ No significant differences in leptin levels were found between MAFLD patients without fibrosis and healthy controls, or between MAFLD patients with and without fibrosis. Interestingly, a two-sample Mendelian randomization study provided strong evidence suggesting that elevated leptin levels may be causally linked to a reduced risk of MAFLD, indicating a potential protective effect.⁸⁸

Taken together, these findings emphasize the importance of considering weight, sex, age, and other individual factors when interpreting leptin levels in MAFLD. A key unresolved question is whether fluctuations in circulating leptin accurately reflect changes in hepatic leptin levels. Subtle differences in liver leptin content could influence hepatic inflammation and fibrosis, even if such changes are not mirrored in systemic circulation. However, this relationship has yet to be clearly established.

Leptin as a potential therapeutic target for patients with MAFLD

The ob/ob mouse model carries a mutation in the leptin

gene, resulting in leptin deficiency.²³ This deficiency causes severe obesity, primarily due to hyperphagia (overeating) and, to a lesser extent, reduced energy expenditure and physical activity. These mice also develop metabolic abnormalities, including elevated lipid, glucose, insulin, and hepatic fat levels.^{184,185} Their metabolic and endocrine profiles closely resemble those of humans with congenital leptin deficiency. Notably, leptin administration corrects all metabolic and endocrine abnormalities in these animals. In contrast, the *ap2-nSREBP-1c* mouse model, which exhibits impaired adipocyte differentiation and reduced white adipose tissue mass, is commonly used to study lipodystrophy.¹⁸⁶ These mice develop hyperglycemia, hyperinsulinemia, mild hyperphagia, and hepatic steatosis. Leptin treatment reduces appetite, body weight, and liver steatosis while improving glucose levels and insulin sensitivity—benefits that occur independently of caloric restriction.¹⁸⁶ Conversely, diet-induced obesity models—characterized by high body fat and elevated circulating leptin—exhibit minimal or no weight loss even when administered high doses of leptin.^{187,188} Collectively, these findings suggest that leptin therapy is more effective in conditions of leptin deficiency—such as congenital leptin deficiency, lipodystrophies, or fasting-induced hypoleptinemia—than in conditions associated with hyperleptinemia.¹⁸⁵

In clinical studies, an open-label trial of patients with relative leptin deficiency and MASH, one year of metreleptin treatment improved liver fat content and MASH scores in five of seven patients with available liver biopsies, with additional improvements in inflammation, ballooning, and fibrosis in some cases.¹⁸⁹ Similarly, Akinci *et al.* reported that metreleptin significantly reduced overall MASH scores in obese adults with relative leptin deficiency.¹⁹⁰ In a placebo-controlled crossover study, a single leptin dose administered to lean individuals post-fasting enhanced VLDL-TC export and may have prevented the hepatic fat accumulation typically associated with fasting.⁹⁸ Improved insulin sensitivity and appetite suppression could reduce the flux of free fatty acids and carbohydrates to the liver, thereby decreasing de novo lipogenesis and hepatic fat accumulation.⁸² However, despite these promising findings, existing studies are limited by small sample sizes. To date, no clinical trial has demonstrated fibrosis improvement without worsening MASH or MASH resolution without increasing fibrosis. Even in insulin-resistant patients, insulin therapy can effectively reduce blood glucose, suggesting that pharmacological doses of leptin might promote weight loss and improve liver fat content. Yet randomized controlled trials using pegylated recombinant human leptin or metreleptin at various doses have not shown significant benefits in energy expenditure, body composition, weight loss, adrenal hormones, sympathetic nervous system activity, lipid metabolism, or macronutrient utilization.^{191–195} Strategies to overcome leptin resistance—such as leptin sensitization or co-treatment with amylin analogs—have been proposed, but to date, they have not produced significant advances in common obesity patients with relatively low leptin levels.

Future perspectives

Numerous studies and clinical cases have demonstrated that MAFLD imposes substantial long-term health, social, and economic burdens, largely due to its high global prevalence.⁵ Without effective policy interventions, these consequences will continue to escalate. Given the increased cardiovascular and mortality risks associated with MAFLD, a multidisciplinary strategy is essential.² There remains an urgent need for non-invasive diagnostic tools to replace liver biopsy, enabling

earlier detection and monitoring in broader populations. Both extrahepatic factors and intrahepatic mechanisms contribute to MAFLD onset and its progression to MASH.¹⁶ Elucidating the underlying pathophysiology and identifying precise molecular targets for diagnosis and treatment remain crucial research priorities.

Leptin, a hormone closely associated with adiposity and IR, has been identified as an independent predictor of MAFLD onset and progression.^{177,180,183} Although leptin exhibits anti-steatotic properties, it is also implicated in hepatic steatosis and the progression of MAFLD to more advanced stages, including MASH and fibrosis.¹⁶ In the early stages of MAFLD, leptin helps regulate glucose homeostasis, reduce hepatic lipid accumulation, and promote lipid oxidation. However, hyperleptinemia can damage pancreatic β -cells and disrupt JAK2/PI3K signaling, contributing to elevated insulin levels and worsening IR.⁸³ This IR, in turn, inhibits lipid oxidation while promoting TC and fatty acid synthesis.^{45,78,84} Leptin can also directly stimulate inflammation by promoting the M1 macrophage phenotype¹²⁰ and increasing pro-inflammatory cytokines such as IL-6, IL-1 β , and TNF- α .¹²² It supports the proliferation of pro-inflammatory CD4⁺ T cells and B cells while inhibiting regulatory T-cell expansion.^{125,126} These lymphocytes release granzymes that generate mitochondrial ROS and activate caspases, ultimately inducing pyroptosis.¹³⁰ Leptin can further promote ROS formation in both phagocytic and non-phagocytic cells, including endothelial cells, cardiomyocytes, and HSCs.^{152–154} The effects of leptin on cirrhosis and HCC in MAFLD remain poorly defined, and its role in other cirrhosis etiologies is still debated.²⁰ Nonetheless, substantial evidence suggests a pro-tumorigenic role for leptin in HCC associated with liver diseases unrelated to MAFLD.

Leptin therapy has proven effective in individuals with congenital leptin deficiency; however, its broader therapeutic potential remains controversial.¹⁸⁵ This underscores the need for further research into the development of leptin analogs that preserve anti-steatotic properties while avoiding pro-inflammatory and fibrogenic effects. Investigating leptin sensitizers and their use in combination with other therapeutic agents should also be prioritized.⁵⁶

Conclusions

Large-scale observational studies and long-term clinical trials are necessary to establish leptin's efficacy across diverse MAFLD phenotypes. Finally, leptin shows promise as a biomarker for MAFLD diagnosis and monitoring, particularly when combined with glucose, lipid, and metabolic profiling. Future research should explore not only leptin's therapeutic potential but also preventive strategies targeting leptin signaling to mitigate MAFLD development.

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Conflict of interest

The authors have no conflict of interests related to this publication.

Author contributions

Manuscript preparation and drafting (JLW, YX, MLL), study conception and design (JLW, JLL), and drafting of the manuscript (JLW). All authors contributed to the manuscript by revising and editing it for important intellectual content. They gave final approval of the version and agreed to be accountable for all aspects of the work presented here.

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